Invited Review

Neuronal modeling with intracellular calcium signaling

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Abstract: Cytosolic Ca²⁺ ions play an important role in the regulation of numerous aspects of cellular activity in virtually all cell types. There is a complex interaction between the neuronal electrical signals on plasma membrane and the chemical signals of intracellular calcium. Each neuron can be considered as a binary membrane system with plasma membrane and endoplasmic reticulum membrane, and the neuronal endoplasmic reticulum can be regarded as a neuron-within-a-neuron. This review explores the simulation modeling of neuronal dynamics mutually coupled with the intracellular calcium signaling released from endoplasmic reticulum through the inositol 1,4,5-trisphosphate receptor calcium channels. We show that a current trend is to include the intracellular calcium dynamics into the neuronal models, and the frontier of this research is now shifting to spatial neuronal models with diffusing intracellular calcium. It is expected that more important results will be obtained with the neuronal models incorporating the intracellular calcium dynamics, especially the spatial models considering the calcium diffusion both in soma and dendritic branches.

Key words: neuron; channel; calcium signals; noise; network

胞内钙信号作用的神经模型

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摘 要: 细胞溶质内的游离钙离子在许多细胞活动中发挥着重要的作用。对于神经元,细胞膜上的神经电信号和胞内钙离子化学信号之间有着复杂的相互作用,每个神经元都可看作为一个含有细胞膜和内质网膜的双膜系统,而神经细胞的内质网则可视为神经元内的神经元。本综述探讨了神经元膜上神经电信号与内质网钙通道释放的胞内钙信号相耦合的动力学模型。我们认为,计算神经动力学的一个研究方向是考虑包含胞内钙动力学的神经元模拟,而且该研究前沿转向考虑胞内钙波扩散运动的空间神经元模型。包含内质网钙动力学的神经模型,尤其是考虑胞体和树突内钙扩散的空间神经元模型,将加深我们对神经动力学的认识。

关键词: 神经元; 离子通道; 钙信号; 噪音; 网络**中图分类号**: O421; O422; O424; O415.6; O414.22

Cytosolic Ca²⁺ ions play an important role in the regulation of numerous aspects of cellular activity in virtually all cell types [1–5]. This versatility arises through the diverse mechanisms by which Ca²⁺ signals are modulated to act over very different time and distance scales. Neurons use both extracellular and intracellular calcium signals to regulate a great variety of neuronal process-

es, including excitability, associativity, neurotransmitter release, synaptic plasticity, and gene transcription ^[2]. A well established mechanism is the regulation of the external calcium influx through voltage-gated calcium channels or ligand-gated ion channels. For example, at synaptic junctions the voltage-gated channels are used to trigger the release of neurotransmitter and to contrib-

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ute to dendritic action potentials. At postsynaptic sites, the neurotransmitters can induce a calcium influx using ligand-gated channels such as the NMDA receptors [6-8].

Although much is known about the influx regulation of external calcium, there is less study on the mechanisms and roles of the intracellular calcium released from the internal calcium stores, such as the endoplasmic reticulum (ER) in neurons ^[9]. The ER is a continuous network distributed throughout the neuron, including the soma, dendrite and axon, as a high calcium concentration store. Throughout the ER the inositol 1,4,5-trisphosphate receptors (IP₃R) or ryanodine receptors (RyR) are distributed ^[10,11]. These receptor channels are responsible for releasing calcium ions from the ER upon the binding of calcium ions on the receptors.

As a result, there is a mutual interaction between the neuronal electrical signals on plasma membrane and the chemical signals of intracellular calcium. The extracellular Ca²⁺ influx during a neuronal action potential can cause the opening of IP₃R or RyR channels to release calcium ions from ER [10,11]. The changes of calcium concentrations in cytosol or ER will modulate the various calcium activated channels on plasma membrane and affect the neuronal excitability [12]. In recent years, there is an increasing interest for neuron modelers to simulate the interaction among the neuronal electrical signals and intracellular calcium signals. This review explores the simulation modeling of neuronal dynamics modulated by the intracellular calcium released from ER IP₃R channels.

1 The concept of neuron-within-a-neuron

Entry of external Ca²⁺ through voltage-operated or receptor-operated channels plays a major role in neuronal activities, but there also is an important contribution of Ca²⁺ released from the ER. The ER is a continuous membrane network that extends throughout the neuron (Fig. 1). The ER maintains a large concentration gradient of Ca²⁺ that can be released in a regenerative manner, thus allowing information to spread long distances through Ca²⁺ waves. This regenerativity is mediated by either IP₃R or RyR, which displays a process of Ca²⁺ induced Ca²⁺ release. One of the important factors determining the sensitivity of these two Ca²⁺ channels is the content of Ca²⁺ in the ER lumen. During a neuronal spike, the Ca²⁺ that enters from the outside is taken up by the ER, which then becomes primed, i.e., the IP₃Rs

and the RyRs become sufficiently sensitive to release the accumulated Ca²⁺ back into the cytosol. In effect, the ER provides a short-term memory of neural activity by integrating the brief Ca²⁺ pulses associated with each discharge. When the accumulated Ca²⁺ reaches the critical threshold for regenerative release, the large explosive and global release of Ca²⁺ could provide the neuron with information concerning previous levels of neuronal activity.

Indeed, the ER network has many properties resembling those of the plasma membrane. Both systems are highly nonlinear. The ER rapidly integrates large amounts of excitatory and inhibitory inputs and triggers an action potential when the membrane potential declines below a critical threshold. Likewise, the excitability of the ER membrane is highly variable and also has to reach a critical threshold before it will elicit a regenerative Ca²⁺ wave. These two membranes are intimately tied together through a variety of reciprocal interactions to regulate specific neuronal processes such as excitability, associativity, transmitter release, synaptic plasticity, and gene transcription [6–8].

Thus, both structurally and functionally the ER may be considered as 'a neuron-within-a-neuron'. The concept of a neuron-within-a-neuron was first suggested by Berridge in 1998 ^[2]. The basic idea is that neuronal Ca²⁺ signaling depends upon a binary membrane system, i.e. the outer plasma membrane and the internal ER membrane (Fig. 1). The outer plasma membrane integrates external information (e.g. excitatory and inhibitory inputs) and generates fast propagating action potentials using voltage-dependent Na⁺ and Ca²⁺ channels. The ER system monitors internal signals, e.g. Ca²⁺ and the inositol 1,4,5-trisphosphate (IP₃) messengers, and can produce slowly propagating regenerative Ca²⁺ signals using a conduction system based on the IP₃Rs.

2 Release of calcium from ER store

In many cells ER stores are the primary source of Ca²⁺ ions. The IP₃R channel opens upon the binding of both Ca²⁺ ion and IP₃, a messenger generated in response to binding of numerous extracellular ligands to Gq- and tyrosine-kinase-coupled cell-surface receptors ^[10]. Cells have developed an active Ca²⁺ propagation mechanism, involving Ca²⁺-induced Ca²⁺ release, by which diffusing Ca²⁺ ions released from an open channel can bind to nearby IP₃R channels to open them, inducing more ER Ca²⁺ release. This regeneration process will be terminated because high Ca²⁺ concentrations cause

excessive Ca²⁺ binding on IP₃R channel to close the channel. Finally, cytosolic calcium declines as cellular pump process drives Ca²⁺ ions back into ER and out of the cell. Governed by such nonlinear reaction-diffusion processes, complex spatiotemporal waves of Ca²⁺ concentration are generated (Fig. 2).

At rest the cytosolic Ca²⁺ is maintained very low (50–100 nmol/L), but much higher concentrations are generated upon the opening of IP₃Rs. Ca²⁺ ions in the cytosol move by passive diffusion alone. This process

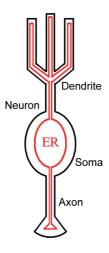


Fig. 1. Binary membrane system of neuronal Ca²⁺ signaling with the outer plasma membrane (black lines) and the internal ER membrane (red lines). The ER is a continuous network that extends to all parts of the neuron, including soma, dendrite and axon, which can be treated as a neuron-within-a-neuron.

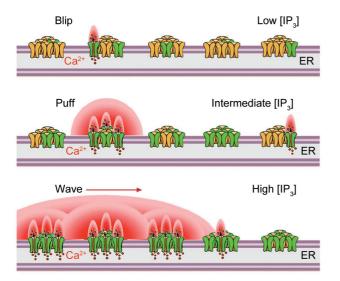


Fig. 2. The hierarchy structure of Ca²⁺ signals release from ER IP₃R channels: blips from single IP₃R, puffs from a cluster of IP₃Rs and global waves spreading through the cell. The figure which was modified from Parker *et al.*, 1996 [19], is kindly provided by Prof. Ian Parker in UC Irvine.

is modified by the presence of endogenous mobile and immobile Ca^{2+} buffers that bind free Ca^{2+} and generally slow down its effective rate of diffusion, steepening Ca^{2+} microdomains around an open Ca^{2+} channel [13]. Thus, the Ca^{2+} concentration near the channel mouth may be 100 µmol/L or more, whereas concentrations as close as one or two µm fall below 1 µmol/L [3, 12, 14]. Ca^{2+} ions therefore have only a restricted "range of action", on the order of 5 µm [15]. However, the positive regulation of IP_3R by Ca^{2+} enables long-range signaling through the generation of actively propagating Ca^{2+} waves which travel at a few dozen µm/s by successive cycles of Ca^{2+} release, diffusion, and dynamics of Ca^{2+} induced Ca^{2+} release [16, 17].

Imaging technology shows that IP₃R channels are distributed on ER membrane in highly localized clusters on hundred-nanometer scale, spaced a few micrometers apart [18-20]. Thus, the intracellular Ca²⁺ signals show a hierarchy structure (Fig. 2) [1, 17, 21, 22]. Weak activation by low IP, concentration evokes localized elevations of cytosolic Ca²⁺ concentration, which arises stochastically and autonomously at discrete release sites. These events are of variable size. The smallest involve Ca2+ flux through single IP3R, and are called blips [23-26]. Larger events (puffs) involve concerted opening of multiple channels clustered at spacing of a few dozen nanometers within individual release sites [18, 19, 24, 25]. These puff events were first observed in Xenopus oocytes [18] and have now been observed in numerous other cell types, e.g. cultured tumor cells, secretory cells, and neurons [1, 27, 28], suggesting they are a ubiquitous feature of IP₃/Ca²⁺ signaling. With higher IP₃ concentration, Ca²⁺ released at one site can trigger Ca²⁺ release at adjacent sites, leading to the generation of Ca²⁺ waves that propagate in a saltatory manner [17, 24, 29].

3 Calcium-activated ion channels

Release of Ca^{2+} from the ER modulates neuronal excitability by altering Ca^{2+} -dependent ion channel currents. The Ca^{2+} -dependent ion channels include the calciumactivated potassium channels $(K_{Ca})^{[3,30]}$, store-operated calcium (SOC) channels $^{[31]}$, calcium-release-activated calcium (CRAC) channels $^{[32]}$, and calcium-dependent chloride channels (Fig. 3). As an example we simply review the K_{Ca} channels $^{[3,30]}$. We discuss the two classes of K_{Ca} channels that have distinct intrinsic affinities for Ca^{2+} , i.e. the voltage- and Ca^{2+} -activated large conductance K^+ channels (BK_{Ca}) and Ca^{2+} -activated small

conductance K⁺ channels (SK_{Ca}).

 BK_{Ca} channels are involved in a diversity of physiological processes ranging from regulation of smooth muscle tone to modulation of neurotransmitter release [33, 34]. BK_{Ca} channels are activated by the cooperative effects of two distinct stimuli, membrane depolarization and cytoplasmic Ca^{2+} . Both stimuli converge allosterically on the gating apparatus of the channels, with increasing Ca^{2+} concentrations shifting the activation curve from highly positive potentials (> 100 mV) into the physiological voltage range [35]. Robust activation of BK_{Ca} channels at membrane potentials around 0 mV requires values for Ca^{2+} concentration > 10 μ mol/L [36], as are known to only occur in the immediate vicinity of active Ca^{2+} sources, particularly Ca_V channels.

According to the distance, there are two classes of BK_{Ca} channels in neurons: those tightly associated with Ca_V channels in Ca^{2+} nanodomains and those located more distantly from Ca^{2+} sources. The macromolecular BK_{Ca} - Ca_V complexes that physically link a Ca^{2+} source and a Ca^{2+} -dependent effector may be in the 10 nm range. Formation of macromolecular complexes between BK_{Ca} and Ca_V channels not only provides a simple mechanism for reliably delivering micromolar Ca^{2+} to BK_{Ca} channels without affecting other Ca^{2+} -dependent signaling processes, but also puts the activity of BK_{Ca} channels under tight control of the Ca_V partner. Distinct from localization in Ca^{2+} nanodomains, some

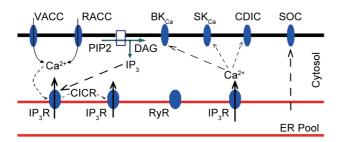


Fig. 3. Neural Ca^{2+} signaling. The extracellular Ca^{2+} ions can enter the neuron through voltage-activated Ca^{2+} channels (VACC) or receptor-activated Ca^{2+} channels (RACC). Metabotropic neurotransmitters stimulate the formation of IP_3 from PIP2, which acts on IP_3R channels to release Ca^{2+} from the ER pool. On ER membrane there are also RyR channels. Both the IP_3Rs and the RyRs are sensitive to Ca^{2+} . The process of Ca^{2+} -induced Ca^{2+} release (CICR) will set up propagated Ca^{2+} waves. The increase of intracellular Ca^{2+} alters Ca^{2+} -dependent ion channels (CDIC) on plasma membrane, including the BK_{Ca} and the SK_{Ca} channels. The store-operated calcium (SOC) channel is controlled by the Ca^{2+} concentration in ER.

 BK_{Ca} channels may also be located more distant from Ca_V channels and be operated by a global increase in Ca^{2+} .

 SK_{Ca} channels are widely expressed in the central nervous system (CNS), where they are important for intrinsic excitability, dendritic integration, and contributing to synaptic plasticity [37–39]. SK_{Ca} channels are gated solely by intracellular Ca^{2+} ions [40, 41]. SK_{Ca} channel activity shows a steep dependence upon Ca^{2+} . Unlike BK_{Ca} channels, where there may be a single coassembled Ca_V channel fueling a single BK_{Ca} channel, SK_{Ca} channels likely are contained within a microdomain with more than a single Ca^{2+} source providing the Ca^{2+} for SK_{Ca} channel activation. Due to their intrinsically higher Ca^{2+} sensitivity, SK_{Ca} channels may be located as far as several dozen nanometers away from the Ca^{2+} sources that are still capable of providing essential Ca^{2+} to open SK_{Ca} channels.

4 Modeling of intracellular calcium dynamics

Since the first observations of Ca²⁺ oscillations, the experimental investigation of their molecular mechanism has been accompanied by numerous modeling approaches. One of the most attractive features about models is their ability to make experimentally testable predictions. That Ca²⁺ oscillations can occur in the presence of a constant level of IP₃ and that the self-amplification of Ca²⁺ release from the ER into the cytoplasm lies at the basis of Ca²⁺ oscillations were for example first predicted theoretically [42]. However, this early model assumed that Ca²⁺ oscillations require the existence of 2 types of pools, some sensitive to IP₃ and others possessing RyR and thereby sensitive to Ca²⁺. This turned out not to be necessary as the IP₃R is itself sensitive to both Ca²⁺ and IP₃.

Models of the IP₃R play a central role not only for understanding channel kinetics, but also as building blocks for constructing larger-scale models of cellular Ca²⁺ signaling. Several IP₃R models [43–49] have been developed to describe experimental data obtained from IP₃R reconstituted in bilayer membranes, with broad application of the De Young-Keizer model [44] in particular. However, there are significant differences in the behavior of reconstituted IP₃Rs in bilayer membrane [50] versus that of IP₃Rs in their native environment of the nuclear envelope [51, 52], and only a few models have incorporated IP₃R data obtained on nucleus membranes [52–54]. Shuai *et al.* [55] have suggested a dynamic

IP₃R model that successfully reproduces data obtained from patch-clamp recordings of nuclear IP₃Rs and is sufficiently simple. The channel model consists of four identical, independent subunits, each of which has an IP₃-binding site, together with one activating and one inactivating Ca²⁺ binding site. A feature of the model is that channel opening may occur if either three or four of the subunits of the tetramer are in an activated state, i.e. binding both IP₃ and the activating Ca²⁺ ion, but not the inhibitory Ca²⁺ ion.

With the IP₃R channel models many computational calcium models have been developed in the past 20 years. They differ by the precise oscillations that they aim to describe, as determined both by the cell type and by the agonist. Models also vary according to the level of description, from the blip modeling with a single IP₃R channel for calcium release [56-60], to the puff modeling with several dozen IP₃R channels distributed in a cluster on ER membrane [59-64], and to the global calcium oscillations and waves [16, 65-71]. For the blip and puff modeling the stochastic channel open/closing dynamics must be considered. The simple models are dimensionless model, and more realistic models consider the diffusion-reaction process of free calcium, stationary and mobile calcium buffers in a two or three-dimensional cell body.

5 Dimensionless somatic neuron models

Sixty years ago, Hodgkin and Huxley proposed a single-compartment somatic neuron model ^[72]. The classic Hodgkin-Huxley model neglects the neuron's spatial structure and focuses entirely on how its various ionic currents contribute to subthreshold behavior and spike generation. These models have led to a quantitative understanding of many dynamical phenomena including phasic spiking, bursting, and spike-frequency adaptation ^[73, 74]. More than 50 years after Hodgkin and Huxley analyzed the squid axon, simple neuron models still offer surprising power on neuronal dynamics.

The dimensionless models to couple the electrical excitability and intracellular calcium oscillation have been discussed several decades ago ^[75]. Chay *et al.* consider in detail different Hodgkin-Huxley-type models with the modulation of intracellular calcium ^[75–79]. One of the models is for R15 Aplysia neurons ^[76, 79]. The R15 neurons model contains a Na⁺ current, an L-type Ca²⁺ current, a cationic nonselective inward current, whose strength depends on the level of luminal Ca²⁺

concentration. With the model it has been shown that the intracellular calcium oscillation can induce the electrical bursting with a much higher spiking frequency. With the bifurcating diagrams, they discussed how neuronal spiking can be transformed to periodic bursting or chaotic behavior with the change of model parameters [77].

Shuai *et al.* propose a sensory cell with the basal membrane contains voltage-sensitive Ca²⁺ channels, voltage-sensitive and Ca²⁺-activated K⁺ channels [80, 81]. While the free intracellular calcium concentration in the sub-membrane space is treated as a dynamic variable, which changes with time following two terms, the influx of extracellular Ca²⁺ ions into the cell through the voltage-sensitive calcium channel and efflux of free intracellular Ca²⁺ ions from the sub-membrane space to the extracellular medium by the pump action and to the intracellular medium by absorption. With the model, the functional differences between A- and B-receptors of weakly electric fish *G. petersii* are explained [80,81].

Kusters *et al.* consider the cell membrane potential depending on the inward rectifier potassium channels, the L-type Ca²⁺ channels, SOC channels, and Ca²⁺ dependent chloride channels ^[82]. The Cl⁻ channel current increases with cytosolic Ca²⁺ concentration, causing a depolarization to the Nernst potential of Cl⁻ ions near –20 mV. The SOC channel current increases when the concentration of Ca²⁺ in the ER decreases ^[83]. The model shows that the coupling of the two oscillation systems, i.e. the periodic firing of membrane action-potential and the periodic oscillation of intracellular Ca²⁺ released from the ER, leads to a rich behavior, including multiple stable and unstable states and hysteresis, in agreement with experimental observations ^[83].

The modeling results also indicate that a high Ca²⁺ buffer concentration can alter the characteristic regular firing of cerebellar granule cells and that a transition to various modes of oscillations occurs, including bursting ^[84]. These simulation results suggest that cytosolic Ca²⁺ buffering capacity can tightly modulate neuronal firing patterns leading to generation of complex patterns and therefore that Ca²⁺-binding proteins may play a critical role in the non-synaptic plasticity and information processing in the CNS.

6 Spatial somatic neuron models

Beyond the dimensionless neuronal models, one can simulate the spatial neuron models. Comparing to the fast spreading of action potential on plasma membrane, the spreading speed of intracellular Ca^{2+} wave is very slow at several dozen μ m/s, thus the spatial models in a two or three-dimensional cell body can only describe the Ca^{2+} diffusion dynamics.

Fletcher and Li consider a single spherical soma model with the ion channels evenly distributed on the cell surface [85]. With the spherical soma the Ca²⁺ dynamics has been simulated in a three-dimensional cell body with two compartments, only the cytosol and ER. Both compartments are assumed to exist at every space point inside the cell, giving rise to a bidomain model. The cytosol and ER are separated by the ER membrane, which contains IP₃R channels, SERCAs, and a nonspecific Ca²⁺ leak. This approach retains a correct account of the amount of Ca²⁺ entry into the cell and generates a realistic Ca²⁺ profile in the whole cell including the shell area. The Ca²⁺ dynamics are governed by the partial and differential equations and boundary conditions. Rapid Ca²⁺ buffering effect is considered in the cytosol and ER also.

At the plasma membrane electrical activity drives Ca²⁺ ions into the cytosol through Ca²⁺ channels, whereas Ca²⁺ ions are extruded by the plasma membrane Ca²⁺ ATPases and Na⁺-Ca²⁺ exchangers. The membrane electrical activity is governed by seven channel currents, including a TTX-sensitive Na⁺ current, an L-type Ca²⁺ current, a delayed rectifier K⁺ current, an inward rectifier K⁺ current, a small-conductance Ca²⁺-activated K⁺ current, a cAMP-activated nonspecific cation current, an SOC current that is inhibited by high Ca²⁺ concentration in ER. Such a model not only can account for the various experimental results, but also predicts a bursting mechanism which has not been reported in experiments [85].

7 Dendritic neuron models

An interesting problem is how the spatial structure of a neuron contributes to its dynamics and function. In order to answer it, one has to design the morphologically realistic models based on anatomical reconstructions. These models extend the cable theory of Rall, who showed mathematically that dendritic voltage attenuation spreads asymmetrically [86]. This phenomenon allows dendrites to compute the direction of synaptic activation patterns, and thus provides a mechanism for motion detection. When voltage-dependent conductances are taken into account, numerical integration

over the spatially discretized dendrite, the compartmental model, is needed to solve the resulting high-dimensional system of equations [87–90]. For complex dendritic trees, more than 1 000 compartments are required to capture the cell's specific electrotonic structure, e.g. the spike back propagation in pyramidal neurons [91].

Combining with Ca²⁺ dynamics, Loewenstein and Sompolinsky show that the temporal integration can be achieved at a single neuron level ^[92]. The calculation and memory of position variables by temporal integration of velocity signals is essential for posture, the vestibule ocular reflex and navigation ^[93]. Integrator neurons exhibit persistent firing at multiple rates, which represent the values of memorized position variables. A widespread hypothesis is that temporal integration is the outcome of reverberating feedback loops within recurrent networks ^[94, 95]. However, Loewenstein and Sompolinsky show that a single neuron can be a neural integrator ^[92].

In the Loewenstein-Sompolinsky model the Ca²⁺ wave-fronts within single neurons can generate graded persistent activity and temporally integrate incoming inputs. The diffusion of Ca²⁺ together with its nonlinear autocatalytic dynamics give rise to wave-fronts of high Ca²⁺ concentrations along dendritic processes of neuron [96]. These fronts propagate at a speed that is proportional to the synaptic input to the cell, resulting in instantaneous front locations that vary in proportion to the temporal integration over previous inputs. Ultimately, information stored in the Ca2+ signals must be decoded by the membrane potential at the axon hillock and communicated by action potentials. In the model, a Ca2+-dependent cationic current has been incorporated into the neuron model. The Ca²⁺-dependent cationic channels are open solely in regions of the dendrite where Ca²⁺ concentration is high. Consequently, the total amount of depolarizing current depends on the location of the Ca²⁺ front along the dendrite. As a result, the Ca²⁺dependent currents translate the location of the fronts into concomitant persistent spiking activity [92].

8 Neuronal network models

The neurons couple to each other to consist complex networks $^{[6-8]}$. There are different coupling mechanisms, including active chemical synaptic interaction, electrotonic coupling through gap junctions, electrical field effects (i.e. ephaptic transmission), and ionic interactions (e.g. increases in the extracellular concentration of K^+).

At the network level much richer dynamics can be found. As the first step one can consider the dimensionless somatic Ca²⁺-neuron networks.

Bondarenko and Chay consider a neuronal network coupled by synaptic connection with intracellular Ca²⁺ dynamics in each dimensionless neuron [97]. The neurons are either excitatory or inhibitory, and so there are three types of couplings: purely excitatory AMPA, purely inhibitory GABA, and mixed connections. These synaptic connections are modified by the intracellular Ca²⁺ of the presynaptic cell, which in turn depends on the Ca2+ ions coming from the Ca2+ channel in the plasma membrane and that released from ER. On the neuronal membrane there are three types of channels, including the voltage-dependent Ca²⁺ channels, the voltage-independent Ca2+ channels, and the delayed-rectifying time-dependent K⁺ channels. The Ca²⁺ concentrations in cytosol and ER are also simulated in the model. The neural network model can produce different types of activities, including spontaneous propagating waves which are observed in the experiments. Such a wave motion can be important for the processes of learning and recognition in neural ensembles.

Loewenstein *et al.* present a dynamic mechanism by which the electrical coupling of identical nonoscillating cells can generate synchronous membrane potential oscillations [98]. They demonstrate this mechanism by constructing a biologically feasible model of electrically coupled cells, characterized by an excitable membrane and intracellular Ca²⁺ dynamics. They show that strong electrical coupling in this network generates multiple oscillatory states with different spatial-temporal patterns. In many studies it usually has been assumed that electrotonic coupling serves as a synchronizing device, or as a fast excitatory pathway. Loewenstein *et al.* suggest that in addition, electrical coupling can serve as a generator of oscillatory activity with the interaction of intracellular Ca²⁺ [98].

9 Noise effects on neuronal dynamics

In the past decades, constructive effects of noise in nonlinear neuronal systems have been investigated extensively in the context of noise-induced transition and noise-induced synchronization [99-103]. Noise-induced transition means that noise may lead to the appearance of new regimes which are not observed in the corresponding noise-free system [99]. An example is the noise-induced spiking or bursting in neural models [101].

Noise-induced synchronization means that a common external noise input to two independent systems could give rise to synchronized motion of both systems [101, 102].

Due to the clustered distribution of IP₃R channels on ER membrane, the IP₃Rs show a strong stochastic channel dynamics, resulting in a strong stochasticity in Ca²⁺ signals. Besides the noisy intracellular Ca²⁺ signals, there are also the noisy currents due to the stochastic dynamics of channels on plasma membrane and the noisy inputs from coupled neurons in the networks. A general question is how the neurons can work with the binary membrane system, either at the single neuron level or at the network level, to exploit these various stochasticities positively and to process neuronal information precisely.

Lang *et al.* discuss the generation and synchronization of bursts in intrinsically spiking neurons due to stimulation with random intracellular Ca²⁺ fluctuations [104]. It is demonstrated that sufficiently strong noise could induce qualitative change in the firing patterns of a single neuron from periodic spiking to bursting modes. Furthermore, it is found that a pair of uncoupled and nonidentical spiking neurons, subjected to a common noise, can exhibit synchronous firing in terms of noise-induced bursting. The synchronization is overall enhanced with the noise intensity increasing, and synchronization transitions are exhibited at intermediate noise levels.

10 Conclusions and future trends

As described in this review, both the plasma membrane and the ER membrane form a binary membrane neuron system. The mutual interaction between the neuronal electrical signals on plasma membrane and the chemical signals of intracellular Ca²⁺ regulates a variety of neuronal processes. The concept of a neuron-within-aneuron becomes all the more interesting because the ER membrane, like the plasma membrane, has both integrative and regenerative properties that could play important roles in neural signaling ^[2].

Since Hodgkin and Huxley proposed the neuron model in 1952 [72], the quantitative neuronal models explain and organize the rapidly growing amount of experimental data, and make lots of testable predictions. As the neuronal models and experiments become more closely interwoven, the more realistic neuronal models are required. Thus, a current trend in neuronal simulations is to include the intracellular Ca²⁺ dynamics into

the models.

In this review we show that in recent years there are an increasing number of neuronal models on the mutual interaction among the neuronal electrical signals and the intracellular Ca^{2+} signals. However, most of these models are dimensionless. The frontier of this interest is now shifting to spatial neuronal models coupled with diffusing Ca^{2+} .

With the spatial models, one can discuss not only the slow Ca^{2+} diffusion dynamics, but also the questions how the hierarchy Ca^{2+} signals, i.e. the local blips and puffs, and global waves, module neuronal activity differently. As reviewed in the paper, the effective coupling range is in nanodomain between BK_{Ca} and Ca_{V} channels, and also can be in microdomain between SK_{Ca} and Ca_{V} channels. In order to simulate these different couplings with different distances, one has to go beyond the dimensionless neuronal models and consider the spatial structure of the neuron.

With the spatial models, it is also possible to explain why both the voltage-gated ion channels on the plasma membrane and the IP₃R channels on the ER membrane have to be at the right place in the right group to couple to each other and to endow individual neurons with their specific character, and how the biophysical properties of channels together with their spatial distribution can define the signalling characteristics of a neuron [105, 106]. It is expected that more important results and conclusions will be obtained with the spatial model to incorporate the Ca²⁺ diffusion in soma and dendritic branches at the single neuron level or at the network level.

REFERENCES

- 1 Bootman MD, Berridge MJ, Lipp P. Cooking with calcium: the recipes for composing global signals from elementary events. Cell 1997; 91: 367–373.
- 2 Berridge MJ. Neuronal calcium signaling. Neuron 1998; 21: 13–26.
- 3 Neher E. Vesicle pools and Ca²⁺ microdomains: new tools for understanding their roles in neurotransmitter release. Neuron 1998; 20: 389–399.
- 4 Berridge MJ, Lipp P, Bootman MD. The versatility and universality of calcium signalling. Nat Rev Mol Cell Biol 2000; 1: 11–21.
- 5 Clapham DE. Calcium signaling. Cell 2007; 131: 1047– 1058.
- 6 Greer PL, Greenberg ME. From synapse to nucleus: Calcium-dependent gene transcription in the control of synapse development and function. Neuron 2008; 59: 846–860.

- 7 Catterall WA, Few AP. Calcium channel regulation and presynaptic plasticity. Neuron 2008; 59: 882–901.
- 8 Higley MJ, Sabatini BL. Calcium signaling in dendrites and spines: practical and functional considerations. Neuron 2008; 59: 902–913.
- 9 Verkhratsky A. Physiology and pathophysiology of the calcium store in the endoplasmic reticulum of neurons. Physiol Rev 2005; 85: 201–279.
- Foskett JK, White C, Cheung KH, Mak DO. Inositol trisphosphate receptor Ca²⁺ release channels. Physiol Rev 2007; 87: 593–658.
- 11 Fill M, Copello JA. Ryanodine receptor calcium release channels. Physiol Rev 2002; 82: 893–922.
- 12 Fakler B, Adelman JP. Control of K_{Ca} channels by calcium nano/microdomains. Neuron 2008; 59: 873–881.
- 13 Pando B, Ponce DS, Mak DO, Pearson JE. Messages diffuse faster than messengers. Proc Natl Acad Sci U S A 2006; 103: 5338–5342.
- 14 Rios E, Stern MD. Calcium in close quarters: microdomain feedback in excitation-contraction coupling and other cell biological phenomena. Annu Rev Biophys Biomol Struct 1997; 26: 47–82.
- 15 Allbritton NL, Meyer T, Stryer L. Range of messenger action of calcium ion and inositol 1,4,5-trisphosphate. Science 1992; 258: 1812–1815.
- Ponce DS, Keizer J, Pearson JE. Fire-diffuse-fire model of dynamics of intracellular calcium waves. Proc Natl Acad Sci U S A 1999; 96: 6060–6063.
- 17 Berridge MJ. Elementary and global aspects of calcium signalling. J Physiol 1997; 499: 291–306.
- 18 Parker I, Yao Y. Regenerative release of calcium from functionally discrete subcellular stores by inositol trisphosphate. Proc R Soc Lond B Biol Sci 1991; 246: 269–274.
- 19 Parker I, Choi J, Yao Y. Elementary events of InsP3-induced Ca²⁺ liberation in *Xenopus* oocytes: hot spots, puffs and blips. Cell Calcium 1996; 20: 105–121.
- 20 Smith IF, Wiltgen SW, Shuai JW, Parker I. Ca²⁺ puffs originate from preestablished clusters of inositol trisphosphate receptors. Sci Signal 2009; 2: ra77.
- 21 Marchant JS, Parker I. Functional interactions in Ca²⁺ signaling over different time and distance scales. J Gen Physiol 2000; 116: 691–696.
- 22 Bootman MD, Lipp P, Berridge MJ. The organisation and functions of local Ca²⁺ signals. J Cell Sci 2001; 114: 2213–2222.
- 23 Sun XP, Callamaras N, Marchant JS, Parker I. A continuum of InsP3-mediated elementary Ca²⁺ signalling events in *Xe-nopus* oocytes. J Physiol 1998; 509: 67–80.
- 24 Rose HJ, Dargan S, Shuai JW, Parker I. 'Trigger' events precede calcium puffs in *Xenopus* oocytes. Biophys J 2006;

- 91: 4024-4032.
- 25 Smith IF, Parker I. Imaging the quantal substructure of single IP₃R channel activity during Ca²⁺ ouffs in intact mammalian cells. Proc Natl Acad Sci U S A 2009; 106: 6404–6409.
- 26 Parker I, Smith IF. Recording single-channel activity of inositol trisphosphate receptors with a microscope, not a patch clamp. J Gen Physiol 2010; 136: 119–127.
- 27 Horne JH, Meyer T. Elementary calcium-release units induced by inositol trisphosphate. Science 1997; 276: 1690– 1693.
- 28 Kidd JF, Fogarty KE, Tuft RA, Thorn P. The role of Ca²⁺ feedback in shaping InsP3-evoked Ca²⁺ signals in mouse pancreatic acinar cells. J Physio 1999; 520: 187–201.
- 29 Marchant JS, Parker I. Role of elementary Ca²⁺ puffs in generating repetitive Ca²⁺ oscillations. EMBO J 2001; 20: 65–76.
- 30 Augustine GJ, Santamaria F, Tanaka K. Local calcium signaling in neurons. Neuron 2003; 40: 331–346.
- Putney JW Jr. Store-operated calcium channels: how do we measure them, why do we care? Sci STKE 2004; 2004 (243): pe37.
- 32 Zhang SL, Yu Y, Roos J, Kozak JA, Deerinck TJ, Ellisman MH, Stauderman KA, Cahalan MD. STIM1 is a Ca²⁺ sensor that activates CRAC channels and migrates from the Ca²⁺ store to the plasma membrane. Nature 2005; 437: 902–905
- 33 Latorre R, Brauchi S. Large conductance Ca²⁺-activated K⁺ (BK) channel: activation by Ca²⁺ and voltage. Biol Res 2006; 39: 385–401.
- 34 Sausbier U, Sausbier M, Sailer CA, Arntz C, Knaus HG, Neuhuber W, Ruth P. Ca²⁺-activated K⁺ channels of the BK-type in the mouse brain. Histochem Cell Biol 2005; 125: 725–741.
- 35 Cui J, Cox DH, Aldrich RW. Intrinsic voltage dependence and Ca²⁺ regulation of mslo large conductance Ca²⁺-activated K⁺ channels. J Gen Physiol 1997; 109: 647–673.
- 36 Brenner R, Jegla TJ, Wickenden A, Liu Y, Aldrich RW. Cloning and functional characterization of novel large conductance calcium-activated potassium channel beta subunits, hKCNMB3 and hKCNMB4. J Biol Chem 2000; 275: 6453–6461.
- 37 Cai X, Liang CW, Muralidharan S, Kao JP, Tang CM, Thompson SM. Unique roles of SK and Kv4.2 potassium channels in dendritic integration. Neuron 2004; 44: 351– 364.
- 38 Ngo-Anh TJ, Bloodgood BL, Lin M, Sabatini BL, Maylie J, Adelman JP. SK channels and NMDA receptors form a Ca²⁺-mediated feedback loop in dendritic spines. Nat Neurosci 2005; 8: 642–649.

- 39 Lin MT, Lujan R, Watanabe M, Adelman JP, Maylie J. SK2 channel plasticity contributes to LTP at Schaffer collateral-CA1 synapses. Nat Neurosci 2008; 11: 170–177.
- 40 Kohler M, Hirschberg B, Bond CT, Kinzie JM, Marrion NV, Maylie J, Adelman JP. Small-conductance, calciumactivated potassium channels from mammalian brain. Science 1996; 273: 1709–1714.
- 41 Hirschberg B, Maylie J, Adelman JP, Marrion NV. Gating of recombinant small-conductance Ca²⁺-activated K⁺ channels by calcium. J Gen Physiol 1998; 111: 565–581.
- 42 Goldbeter A, Dupont G, Berridge MJ. Minimal model for signal-induced Ca²⁺ oscillations and for their frequency encoding through protein phosphorylation. Proc Natl Acad Sci U S A 1990; 87: 1461–1465.
- 43 Atri A, Amundson J, Clapham D, Sneyd J. A single-pool model for intracellular calcium oscillations and waves in the *Xenopus laevis* oocyte. Biophys J 1993; 65: 1727–1739
- 44 De Young GW, Keizer J. A single-pool inositol 1,4,5-trisphosphate-receptor-based model for agonist-stimulated oscillations in Ca²⁺ concentration. Proc Natl Acad Sci U S A 1992; 89: 9895–9899.
- 45 Kaftan EJ, Ehrlich BE, Watras J. Inositol 1,4,5-trisphosphate (InsP3) and calcium interact to increase the dynamic range of InsP3 receptor-dependent calcium signaling. J Gen Physiol 1997; 110: 529–538.
- 46 Sneyd J, Dufour JF. A dynamic model of the type-2 inositol trisphosphate receptor. Proc Natl Acad Sci U S A 2002; 99: 2398–2403.
- 47 Swillens S, Champeil P, Combettes L, Dupont G. Stochastic simulation of a single inositol 1,4,5-trisphosphate-sensitive Ca²⁺ channel reveals repetitive openings during blip-like Ca²⁺ transients. Cell Calcium 1998; 23: 291–302.
- 48 Fraiman D, Dawson SP. A model of the IP3 receptor with a luminal calcium binding site: stochastic simulations and analysis. Cell Calcium 2004; 35: 403–413.
- 49 Sneyd J, Falcke M. Models of the inositol trisphosphate receptor. Prog Biophys Mol Biol 2005; 89: 207–245.
- 50 Bezprozvanny I, Watras J, Ehrlich BE. Bell-shaped calcium-response curves of Ins(1,4,5)P3- and calcium-gated channels from endoplasmic reticulum of cerebellum. Nature 1991; 351: 751–754.
- 51 Mak DO, McBride S, Foskett JK. Inositol 1,4,5-trisphosphate activation of inositol trisphosphate receptor Ca²⁺ channel by ligand tuning of Ca²⁺ inhibition. Proc Natl Acad Sci U S A 1998; 95: 15821–15825.
- 52 Mak DO, McBride SM, Foskett JK. Spontaneous channel activity of the inositol 1,4,5-trisphosphate (InsP3) receptor (InsP3R). Application of allosteric modeling to calcium and InsP3 regulation of InsP3R single-channel gating. J

- Gen Physiol 2003; 122: 583-603.
- Baran I. Integrated luminal and cytosolic aspects of the calcium release control. Biophys J 2003; 84: 1470–1485.
- 54 Shuai JW, Yang D, Pearson JE, Ruediger S. An investigation of models of the IP₃R channel in *Xenopus* oocyte. Chaos 2009; 19: 037105-1–037105-11.
- 55 Shuai JW, Pearson JE, Foskett JK, Mak DO, Parker I. A kinetic model of single and clustered IP3 receptors in the absence of Ca²⁺ feedback. Biophys J 2007; 93: 1151–1162.
- 56 Shuai JW, Parker I. Optical single-channel recording by imaging Ca²⁺ flux through individual ion channels: theoretical considerations and limits to resolution. Cell Calcium 2005; 37: 283–299.
- 57 Ruediger S, Shuai JW, Huisinga W, Nagaiah C, Warnecke G, Parker I, Falcke M. Hybrid stochastic and deterministic simulations of calcium blips. Biophys J 2007; 93: 1847–1857.
- 58 Shuai JW, Pearson JE, Parker I. Modeling Ca²⁺ feedback on a single inositol 1,4,5-trisphosphate receptor and its modulation by Ca²⁺ buffers. Biophys J 2008; 95: 3738–3752.
- 59 Swillens S, Dupont G, Combettes L, Champeil P. From Ca²⁺ blips to Ca²⁺ puffs: theoretical analysis of the requirements for interchannel communication. Proc Natl Acad Sci U S A 1999; 96: 13750–13755.
- 60 Ruediger S, Nagaiah C, Warnecke G, Shuai JW. Ca²⁺ domains around single and clustered IP3 receptors and their modulation by buffers. Biophys J 2010; 99: 3–12.
- 61 Shuai JW, Jung P. Stochastic properties of Ca²⁺ release of inositol 1,4,5-trisphosphate receptor clusters. Biophys J 2002; 83: 87–97.
- 62 Shuai JW, Jung P. Optimal intracellular calcium signaling. Phys Rev Lett 2002; 88: 068102-1–068102-4.
- 63 Shuai JW, Rose HJ, Parker I. The number and spatial distribution of IP3 receptors underlying calcium puffs in *Xenopus* oocytes. Biophys J 2006; 91: 4033–4044.
- 64 Ruediger S, Shuai JW, Sokolov IM. Law of mass action, detailed balance, the modeling of calcium puffs. Phys Rev Lett 2010; 105: 048103-1–048103-4.
- 65 Shuai JW, Huang YD, Ruediger S. Puff-wave transition in an inhomogeneous model for calcium signals. Phys Rev E 2010; 81: 041904-1–041904-8.
- 66 Swillens S, Combettes L, Champeil P. Transient inositol 1,4,5-trisphosphate-induced Ca²⁺ release: A model based on regulatory Ca²⁺-binding sites along the permeation pathway. Proc Natl Acad Sci U S A 1994; 91: 10074–10078.
- 67 Dupont G, Swillens S. Quantal release, incremental detection, and long-period Ca²⁺ oscillations in a model based on regulatory Ca²⁺-binding sites along the permeation pathway. Biophys J 1996; 71: 1714–1722.

- 68 Falcke M, Tsimring L, Levine H. Stochastic spreading of intracellular Ca²⁺ release. Phys Rev E Stat Phys Plasmas Fluids Relat Interdiscip Topics 2000; 62: 2636–2643.
- 69 Pando B, Pearson JE, Ponce SD. Sheet excitability and nonlinear wave propagation. Phys Rev Lett 2003; 91: 258101-1–258101-4.
- 70 Shuai JW, Jung P. Optimal ion channel clustering for intracellular calcium signaling. Proc Natl Acad Sci U S A 2003; 100: 506–510.
- 71 Shuai JW, Jung P. Selection of intracellular calcium patterns in a model with clustered Ca²⁺ release channels. Phys Rev E Stat Nonlin Soft Matter Phys 2003; 67: 031905.
- 72 Hodgkin AL, Huxley AF. A quantitative description of membrane current and its application to conduction and excitation in nerve. J Physiol 1952; 117: 500–544.
- 73 Koch C. Biophysics of Computation: Information Processing in Single Neurons. New York: Oxford University Press, 1998
- 74 Rinzel J, Ermentrout GB. In: Methods in Neuronal Modeling: From Synapses to Networks. Koch C, Segev I. Eds. Cambridge: MIT Press, 1989, 135–169.
- 75 Chay TR. The effect of inactivation of calcium channels by intracellular Ca^{2+} ions in the bursting pancreatic β -cells. Cell Biophys 1987; 11: 77–90.
- Chay TR. Electrical bursting and intracellular Ca²⁺ oscillations in excitable cell models. Biol Cybern 1990; 63: 15–23.
- 77 Chay TR, Fan YS, Lee YS. Bursting, spiking, chaos, fractals, and universality in biological rhythms. Int J Bifurcat Chaos 1995; 5: 595–635.
- 78 Chay TR. Modeling slowly bursting neurons via calcium store and voltage-independent calcium current. Neural Comput 1996; 8: 951–978
- 79 Chay TR. Electrical bursting and luminal calcium oscillation in excitable cell models. Biol Cybern 1996; 75: 419–431.
- 80 Shuai JW, Kashimori Y, Kambara T. Electroreceptor model of the weakly electric fish *Gnathonemus petersii*: I. The model and the origin of differences between A- and B-receptors. Biophys J 1998; 75: 1712–1726.
- 81 Shuai JW, Kashimori Y, Kambara T, Emde G. Eectroreceptor model of weakly electric fish *Gnathonemus petersii*: II. Cellular origin of inverse waveform tuning. Biophys J 1999; 76: 3012–3025.
- Kusters JM, Cortes JM, van Meerwijk WP, Ypey DL, Theuvenet AP, Gielen CC. Hysteresis and bistability in a realistic cell model for calcium oscillations and action potential firing. Phys Rev Lett 2007; 98: 098107-1–098107-4.
- 83 Kusters JM, Dernison MM, van Meerwijk WP, Ypey DL, Theuvenet AP, Gielen CC. Stabilizing role of calcium

- store-dependent plasma membrane calcium channels in action-potential firing and intracellular calcium oscillations. Biophys J 2005; 89: 3741–3756.
- 84 Roussela C, Erneuxb T, Schiffmanna SN, Galla D. Modulation of neuronal excitability by intracellular calcium buffering: From spiking to bursting. Cell Calcium 2006; 39: 455–466.
- 85 Fletchera PA, Li YX. An integrated model of electrical spiking, bursting, and Ca²⁺ oscillations in GnRH neurons. Biophys J 2009; 96: 4514–4524.
- 86 Rall W. In: Neural Theory and Modeling. Reiss RF, Ed. Stanford: Stanford University Press, 1964, 73C97.
- 87 Traub RD, Wong RK, Miles R, Michelson H. A model of a CA3 hippocampal pyramidal neuron incorporating voltageclamp data on intrinsic conductances. J Neurophysiol 1991; 66: 635–650.
- 88 Traub RD, Jefferys JG, Miles R, Whittington MA, Tóth K. A branching dendritic model of a rodent CA3 pyramidal neuron. J Physiol 1994; 481: 79–95.
- 89 Shuai JW, Bikson M, Hahn PJ, Lian J, Durand DM. Ionic mechanisms underlying spontaneous CA1 neuronal firing in Ca²⁺-free solution. Biophys J 2003; 84: 2099–2111.
- 90 Saraga F, Wu CP, Zhang L, Skinner FK. Active dendrites and spike propagation in multicompartment models of oriens-lacunosum/moleculare hippocampal interneurons. J Physiol 2003; 552: 673–689.
- 91 Golding NL, Kath WL, Spruston N. Dichotomy of actionpotential backpropagation in CA1 pyramidal neuron dendrites. J Neurophysiol 2001; 86: 2998–3010.
- 92 Loewenstein Y, Sompolinsky H. Temporal integration by calcium dynamics in a model neuron. Nat Neurosci 2003; 6: 961–967.
- 93 Aksay E, Gamkrelidze G, Seung HS, Baker R, Tank DW. In vivo intracellular recording and perturbation of persistent activity in a neural integrator. Nat Neurosci 2001; 4: 184–193.
- 94 Seung HS, Lee DD, Reis BY, Tank DW. Stability of the

- memory of eye position in a recurrent network of conductance-based model neurons. Neuron 2000; 26: 259–271.
- 95 Koulakov AA, Raghavachari S, Kepecs A, Lisman JE. Model for a robust neural integrator. Nat Neurosci 2002; 5: 775–782.
- 96 Feller MB, Wellis DP, Stellwagen D, Werblin FS, Shatz CJ. Requirement for cholinergic synaptic transmission in the propagation of spontaneous retinal waves. Science 1996; 272: 1182–1187.
- 97 Bondarenko VE, Chay TR. The role of AMPA, GABA, [Ca²⁺]_i, and calcium stores in propagating waves in neuronal networks. Neurocomputing 2000; 32–33: 291–298.
- 98 Loewenstein Y, Yarom Y, Sompolinsky H. The generation of oscillations in networks of electrically coupled cells. Proc Natl Acad Sci U S A 2001; 98: 8095–8100.
- 99 Gammaitoni L, Hanggi P, Jung P, Marchesoni F. Stochastic resonance. Rev Mod Phys 1998; 70: 223–287.
- 100 Jung P, Shuai JW. Optimal sizes of ion channel clusters. Europhys Lett 2001; 56: 29.
- 101 Lindner B, Garca-Ojalvo J, Neiman A, Schimansky-Geier L. Effects of noise in excitable systems. Phys Rep 2004; 392: 321–424.
- 102 Neiman A, Russell D. Synchronization of noise-induced bursts in noncoupled sensory neurons. Phys Rev Lett 2002; 88: 138103-1–138103-4.
- 103 Shuai JW, Jung P. Entropically enhanced excitability in small systems. Phy Rev Lett 2005; 95: 114501-1–114501-4.
- 104 Lang X, Lu Q, Kurths J. Phase synchronization in noisedriven bursting neurons. Phys Rev E 2010; 82: 021909-1– 021909-7.
- 105 Lai HC, Jan LY. The distribution and targeting of neuronal voltage-gated ion channels. Nat Rev Neurosci 2006; 7: 548–562.
- 106 Kole MH, Ilschner SU, Kampa BM, Williams SR, Ruben PC, Stuart GJ. Action potential generation requires a high sodium channel density in the axon initial segment. Nat Neurosci 2008; 11: 178–186.

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